

Microbiological Indicators of Patients with Confirmed Sars-Cov-2 - Infection

1. Oblokulov Abdurashid Raximovich
2. Jalilova Aziza Sadilloevna
3. Mukhtorova Shokhida Abdulloyevna
4. Raximov Farrux Farxodovich

Received 26th Jan 2022,
Accepted 19th Feb 2022,
Online 28th Mar 2022

^{1,2,3,4} Bukhara State Medical Institute
named after Abu Ali ibn Sino
a.oblokulov59@gmail.com

Annotation: The article presents a bacteriological analysis of patients admitted to the Bukhara Regional Infectious Diseases Hospital from March 16, 2020 to February 02, 2022 with a diagnosis of COVID-19 coronavirus infection and confirmed SARS-CoV-2 infection (positive real-time RT-PCR, typical for SARS-CoV-2) Results are shown from fecal samples taken from patients Pr. Vulgaris, Pr. Mirobllis, Kl.phevmoniae, Enterobacter hafniae, from sputum samples of St. Pneumoniae, Can. viridans; S.epidermis, S.aureus were isolated from a blood sample. Most of the isolated microorganisms were sensitive to levofloxacin, amikacin, ciprofloxacin and cefoperazone sulbactam.

Keywords: antibiotic resistance; COVID-19; pneumonia; SARS-CoV-2; sputum culture; blood culture.

Relevance of the topic

In December 2019, several cases of severe pneumonia of unknown origin occurred in Wuhan, China [1], later diagnosed as coronavirus 2019 (COVID-19), whose etiologic agent was SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2). It belongs to the family β of the family Coronaviridae [2, 3]. On March 11, 2020, the World Health Organization (WHO) declared the disease a pandemic. Bacterial superinfection and mortality from SARS-CoV-2 are significantly higher than from any other common respiratory viral syndrome [5, 6]. As of March 14, 2022, 456,797,217 cases of COVID-19 have been confirmed worldwide, including 6,043,094 deaths. (<https://www.who.int/>).

Superinfection with SARS-CoV-2 by other microorganisms, especially bacteria and fungi, is a critical factor in the development of COVID-19, complicating diagnosis, treatment, and prognosis [7,8]. Bacterial superinfection in inpatients with COVID-19 is associated with disease progression and prognosis. This condition increases hospitalization, antibiotic treatment, and mortality in intensive care units [9,11,12].

In patients with confirmed coronavirus infection, antimicrobial therapy plays an important role in the treatment of suspected or confirmed bacterial respiratory infection. As a rule, this kind of therapy is empirical in nature or intended for the treatment of nosocomial infections acquired during hospitalization in hospitalized patients with pneumonia. Patients may also suffer from secondary infections not related to the respiratory tract, such as urinary tract or bloodstream infections [13,14,15].

It is known that the course of respiratory viral infections is often associated with the addition of severe bacterial and fungal infections [16,17,18]. Many researchers note an increase in the development of secondary bacterial infections caused by *Streptococcus pneumoniae*, *Staphylococcus aureus*, *Haemophilus influenzae* and *Aspergillus* sp. However, such data on the prevalence and severity of secondary bacterial infections in hospitalized patients with COVID-19 are currently relatively rare.

One of the possible solutions to justify the prescription of antibacterial drugs for COVID-19 is the use of procalcitonin, a specific biomarker of bacterial infection [6,21]. Procalcitonin has been shown to maintain a distinction between bacterial and viral infection and support early discontinuation of antibiotics without affecting patient mortality in confirmed bacterial infection [22,23]. The use of COVID-19 procalcitonin has been reported and may be an important tool to help reduce the use of antimicrobials [24].

The aim of the study was to determine the microbial landscape and antibiotic susceptibility of the main pathogens isolated from patients with COVID-19.

Materials and methods

3467 patients were diagnosed with COVID-19 coronavirus infection from March 16, 2020 to February 2, 2022. Bacteriological studies were performed on 1169 samples (sputum, feces, blood) taken from them.

Sputum specimens include Sabouraud Dextrose Agar, Endo Agar, Blood Agar, Sari Salt Mannitol Cultures, Salmonella-Shigella Agar (SS Agar), Bismuth Sulphite Agar, Kligler Medium (Kligler Iron Agar), nutrient agar (Nutrient Agar). For bacteriological examination of a blood sample, glucose-Sabouraud-Agar (Sabouraud Dextrose Agar), a blood-agar medium, was used.

All patients with COVID-19 included in this study were diagnosed in accordance with the recommendations for the diagnosis and treatment of pneumonia caused by a new coronavirus infection. All patients had laboratory-confirmed SARS-CoV-2 infection (real-time RT-PCR positive, typical of SARS-CoV-2). Complete blood elements were detected in blood samples by determining the number of leukocytes (WBC), lymphocytes (LYM), mononuclear cells (MONO), neutrophils (NEU). Blood biochemical parameters: aspartate aminotransferase (AST), alanine aminotransferase (ALT), glucose (GLU), urea, creatinine, and C-reactive protein (CRO) were determined using an automatic biochemical analyzer MINDRAY BS-30 (China).

Results and their discussion

As of March 16, 2020, bacteriological studies in 1169 samples taken from 3467 patients hospitalized with COVID-19 coronavirus infection showed positive results. Bacteriological culture of feces was obtained in 928 patients, while a positive result was obtained in 556 (59.9%) patients. Sectional analysis of isolated microorganisms showed that 38.3% of *Pr. Vulgaris*, 32.6% *Pr. Miroblis*, *Kl.pneumoniae* were found in 2.3%, *Enterobacter hafniae* in 26.8% (Fig. 1).

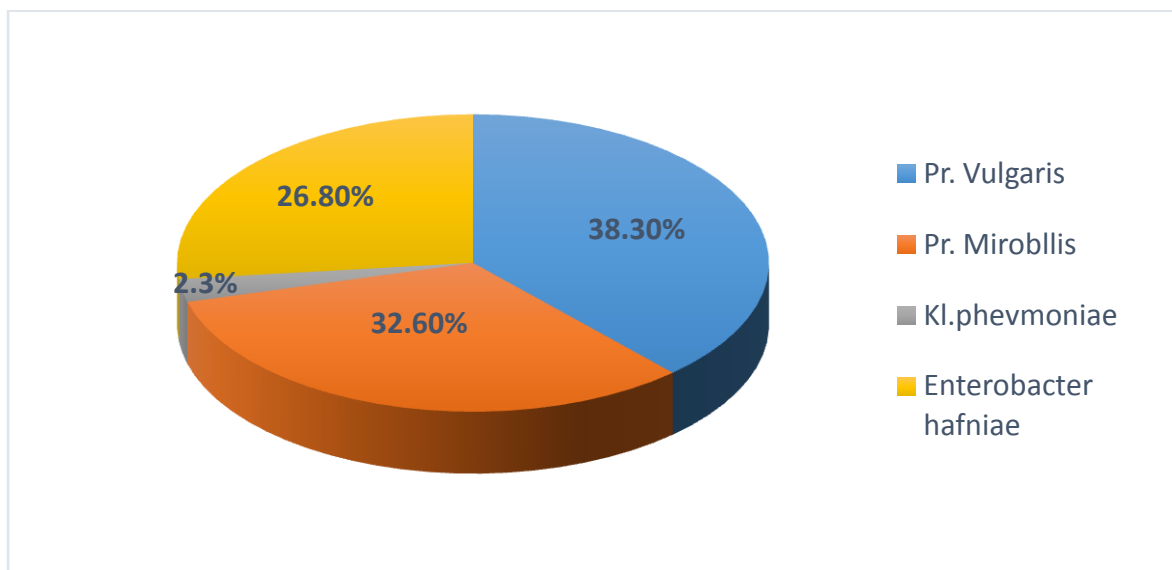


Figure 1. Data of bacteriological inoculation of stool samples

Sputum of 3476 patients was obtained for bacteriological culture, a positive result was obtained in 264 (7.6%). When studying sections of cultures of microorganisms, 61.0% were infected with *S. aureus*, *St. John's wort*. Pneumoniae was found in 5.3% and *Can. viridans* in 33.7% (Fig. 2). In 2299 (66.1%) controlled patients, a bacteriological blood test was performed. Microorganisms multiplied in 15.1% of them. The analysis showed that 71.1% of them were *S. epidermis* and 28.9% *S. aureus*.

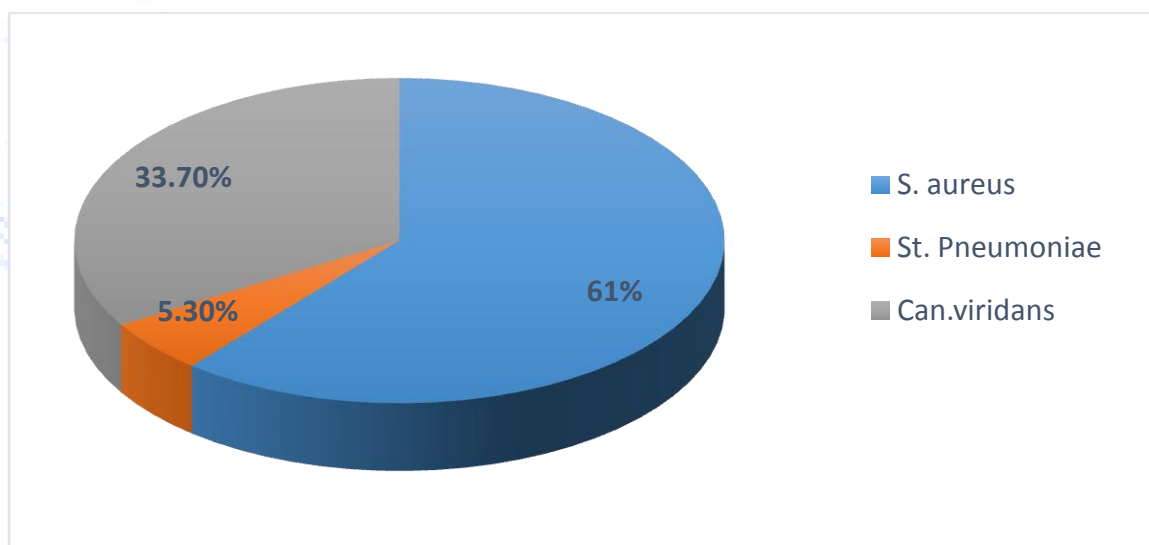


Figure 2. Data of bacteriological inoculation of sputum samples

Of the pathogens identified above, 613 were tested for antibiotic susceptibility. Several antibiotic discs were used to determine the antibiotic susceptibility of the pathogens listed above, and the number and percentage of the most susceptible to antibiotics during the study were as follows.

217 (35.4%) isolated microorganisms were susceptible to levofloxacin, 174 (28.4%) to amikacin, 122 (19.9%) to ciprofloxacin, 61 (9.9%) to cefaperazone sulbactam, 14 (2.4%) % to cefepime and 12 (2.4%) to cefepime. 12 (1.9%) to ceftriaxone. Low sensitivity to cefazidin, cefazidime, gentamicin, chloramphenicol was noted. Sensitivity to semi-synthetic penicillins (benzylpenicillin, ampicillin) has not been identified.

Thus, only 35.5% of the biological extracts obtained for the purpose of basic treatment with antibacterial drugs were microorganisms. Most of them were hypersensitive to levofloxacin, amikacin, ciprofloxacin (Fig. 3).

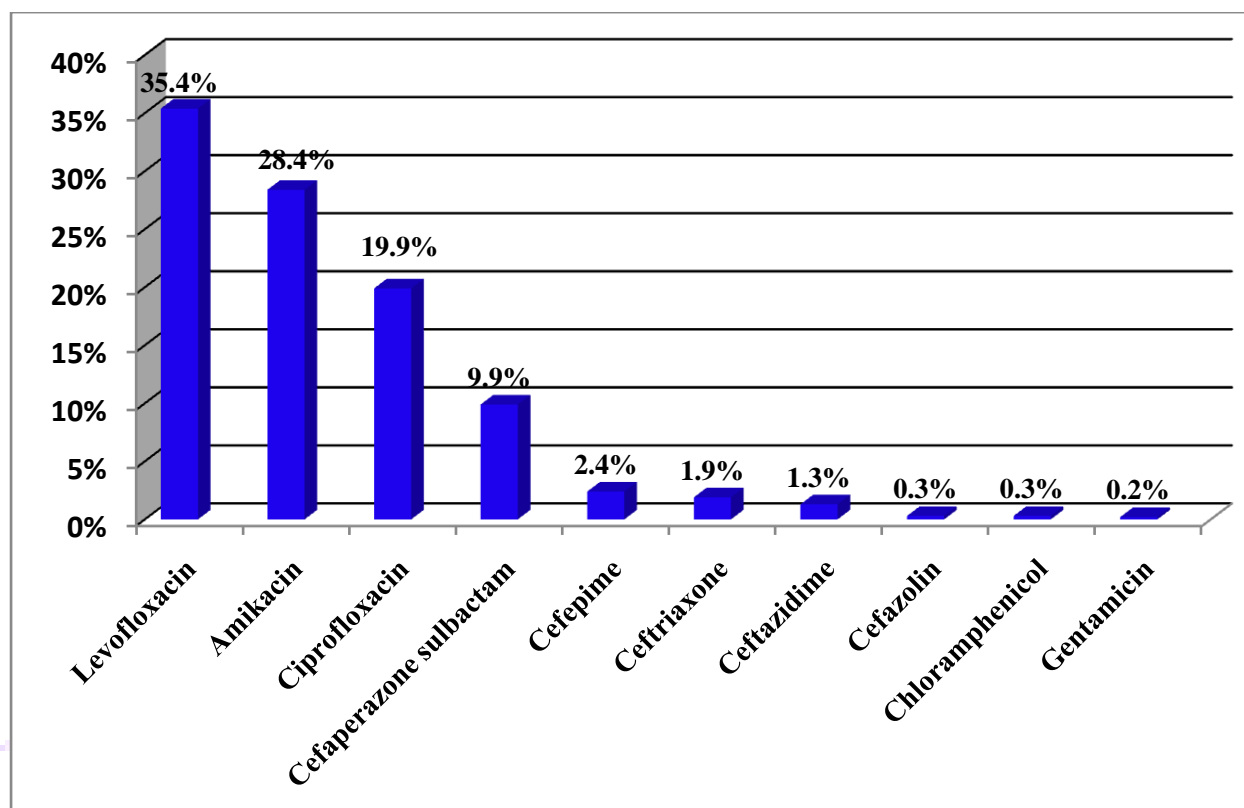


Figure 3. The sensitivity of microorganisms to antibacterial drugs

The microorganisms found in our study were in some respects similar to other published data [25], differing from similar data from studies in China in that pneumococci and *Haemophilus influenza* were the main pathogens [26].

Liu H.H. et al. according to the data, the results of 253 sputum samples did not show acute bacterial or fungal infection in 73 (45%) of 165 people, which were usually collected within 24 hours after intubation. The number of potential pathogens increased 1 week or less after intubation in 72 (64.9%) of 111 cases, indicating late pneumonia in 70.8% and colonization in 29.2%. In 12 of these evening specimens (10.8% of the total), antimicrobial resistance, mainly *Pseudomonas*, *Enterobacter* or *Staphylococcus aureus*, worsened after intubation [27]. In our study, bacteriological culture analysis of sputum samples from patients showed a positive result in 264 (7.6%), of which 61.0% were found to have *S. aureus*, *St. John's wort*. *Pneumoniae* was found in 5.3% and *Can. viridans* in 33.7%. Most of the isolated microorganisms were sensitive to levofloxacin, amikacin, ciprofloxacin and cefoperazone sulbactam.

Conclusion:

1. Most hospitalized patients have co- or COVID-19. advanced forms of superinfection.
2. All patients diagnosed with *can. viridans* received one or another antibacterial drug on an outpatient basis.
3. The isolation of microorganisms from samples taken from the patient substantiates the forms of the disease (pneumonia, gastrointestinal, septic).

4. Most of the isolated microorganisms were sensitive to antibacterial drugs such as levofloxacin, amikacin, ciprofloxacin, cefaperezone, sulbactam.

References

1. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020; 395:497–506.
2. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med* 2020; 382:727–33.
3. Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 2020; 579:270–3.
4. WHO. Coronavirus disease 2019 (COVID-19)—situation report—51. 2020. https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200311-sitrep-51-covid-19.pdf?sfvrsn=1ba62e57_10.
5. Li R, Pei S, Chen B, Song Y, Zhang T, Yang W, et al. Substantial undocumented infection facilitates the rapid dissemination of novel coronavirus (SARS-CoV-2). *Science* 2020; 368 (6490):489–93.
6. Abdurashid Rahimovich Oblokulov, Zilola Zohirovna Husenova, Maksudjon Muzaffarovich Ergashev. (2021). Procalcitonin as an Indicator of Antibacterial Therapy in Covid-19. *Annals of the Romanian Society for Cell Biology*, Volume 25: Issue 3. 5220–5224.
7. Чориевич, Х.Д. и Фарходович Р.Ф. 2021. Клинико-эпидемиология Характеристика Пациентов С Тяжёлой Пневмонии, Вызванной SARS-COV-2. *ЦЕНТРАЛЬНОАЗИАТСКИЙ ЖУРНАЛ МЕДИЦИНСКИХ И ЕСТЕСТВЕННЫХ НАУК*. (октябрь 2021 г.), 178–182. DOI: <https://doi.org/10.47494/cajmns.vi0.373>.
8. Shen Z, Xiao Y, Kang L, Ma W, Shi L, Zhang L, et al. Genomic diversity of SARS-CoV-2 in Coronavirus Disease 2019 patients. *Clin Infect Dis* 2020;71:713–20.
9. Elmurodova A. A. (2022). “Specific Features of the Hemostatic System in Covid-19”. *CENTRAL ASIAN JOURNAL OF MEDICAL AND NATURAL SCIENCES* 3, 82-85.
10. Oblokulov, A. R., Niyozov, G. E., Elmurodova, A. A., & Orifov, D. U. (2020). CLINICAL CHARACTERISTICS OF PATIENTS WITH COVID-19. *Interdisciplinary Approaches to Medicine*, 1(2), 40-43.
11. Martins-Filho PR, Tavares CSS, Santos VS. Factors associated with mortality in patients with COVID-19. A quantitative evidence synthesis of clinical and laboratory data. *Eur J Intern Med* 2020;76:97–9.
12. Oblokulov A.R., Niyozov G.E. (2020). Clinical and epidemiological characteristics of patients with COVID-19. *International Journal of Pharmaceutical Research*, 12 (4), pp. 3749-3752.
13. Priya Nori et al. Bacterial and fungal coinfections in COVID-19 patients hospitalized during the New York City pandemic surge // *Infect Contr and Hospital Epid*, 2020. 1-5.
14. Niyazov G.E., Oblokulov A.R., Pondina A.I. et al. (2020) Clinical and epidemiological characteristics of COVID-19 patients // *New Day in Medicine*. No4 (32) 110-115 p.
15. Г, Худойдодова С., Фармонова М. В. 2021. «Covid-19 У Детей». *Central Asian Journal of Medical and Natural Sciences*, октябрь, 183-86. <https://doi.org/10.47494/cajmns.vi0.374>.

16. MacIntyre C.R., Chughtai A.A., Barnes M, et al. The role of pneumonia and secondary bacterial infection in fatal and serious outcomes of pandemic influenza a (H1N) // BMC Infect Dis. 2018. 18:637.doi.org/10.1186/s12879-018-3548-0.
17. Р., Облокулов А., Ниезов Г.Э., Ражабов А.Р. (2021). “Covid-Ассоциированная Коагулопатия У Пациентов Новой Коронавирусной Инфекции”. *CENTRAL ASIAN JOURNAL OF MEDICAL AND NATURAL SCIENCES*, October, 124-30. <https://doi.org/10.47494/cajmns.vi0.362>.
18. Cataño-Correa JC, Cardona-Arias JA, Porras Mancilla JP, García MT (2021) Bacterial superinfection in adults with COVID-19 hospitalized in two clinics in Medellín-Colombia, 2020. *PLoS ONE* 16(7): e0254671. <https://doi.org/10.1371/journal.pone.0254671>
19. Mulcany M.E., McLoughlin R.M. Staphylococcus aureus and influenza A virus: patterns and coinfections. *VBio*. 2016. 7:e02068-16;
20. Canning B., Sennyake R.V., Burns D., Moran E, Dedicoat M. Post-influenza aspergillus ventriculitis // *Clin Infect Pract* 2020:100026. doi:10.1016/j.clinpr.2020.100026
21. Meier MA, Branche A, Neeser OL, et al. Procalcitonin-guided antibiotic treatment in patients with positive blood cultures: a patient-level meta-analysis of randomized trials. *Clin Infect Dis* 2019; 69:388–96.
22. Meier MA, Branche A, Neeser OL, et al. Procalcitonin-guided antibiotic treatment in patients with positive blood cultures: a patient-level meta-analysis of randomized trials. *Clin Infect Dis* 2019; 69:388–96.
23. De Jong E, van Oers JA, Beishuizen A, et al. Efficacy and safety of procalcitonin guidance in reducing the duration of antibiotic treatment in critically ill patients: a randomised, controlled, open-label trial. *Lancet Infect Dis* 2016; 16:819–27.
24. Timothy M Rawson, Luke S P Moore, Nina Zhu et al. Bacterial and Fungal Coinfection in Individuals With Coronavirus: A Rapid Review To Support COVID-19 Antimicrobial Prescribing *clinical Infectious Diseases*, Volume 71, Issue 9, 1 November 2020, Pages 2459–2468.
25. Verroken A., Scohy A., Gerard L., et al. Co-infections in COVID-19 critically ill and antibiotic management: A prospective cohort analysis // *Crit Care*. 2020.24:410.
26. Hans H. Liu et al. Bacterial and fungal growth in sputum cultures from 165 COVID-19 pneumonia patients requiring intubation: evidence for antimicrobial resistance development and analysis of risk factors. doi: 10.21203/rs.3.rs-79487/v1.
27. Liu HH, Yaron D, Piraino AS, Kapelusznik L. Bacterial and fungal growth in sputum cultures from 165 COVID-19 pneumonia patients requiring intubation: evidence for antimicrobial resistance development and analysis of risk factors. *Ann Clin Microbiol Antimicrob*. 2021 Sep 25;20 (1):69.
28. Садиллоевна, Ж. А., Саётовна К. Д. (2021). Клиника – Лабораторная Характеристика Пациентов С Covid-19 И Предиктор Антибактериальной Терапии. *ЦЕНТРАЛЬНО-АЗИАТСКИЙ ЖУРНАЛ МЕДИЦИНСКИХ И ЕСТЕСТВЕННЫХ НАУК*, 81-86. <https://doi.org/10.47494/cajmns.vi0.354>